

ABSTRACT

"PLASTINATION AS A CLINICALLY BASED TEACHING AID AT THE UNIVERSITY OF AUCKLAND"

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As a concerted move toward closer integration of the clinical and pre-clinical aspects of the undergraduate medical curriculum at the University of Auckland, the Department of Anatomy has established a formal link with the Department of Radiology resulting in a structured program of clinically based teaching of gross anatomy to second and third year medical students.

As sophisticated diagnostic techniques and methods of treatment have become common place, our teaching program has been tailored to accommodate a greater degree of case based learning within the undergraduate course.

Dissecting room demonstration is provided by Radiology, Pathology, Ophthalmology and Surgical registrars, with a number of clinical procedures, pathological observations, and diagnostic methods employed during the routine dissection of the cadaver.

Through use of a number of plastination techniques, the learning process is enhanced and aided on a number of levels. The E-12 epoxy method for producing M.R.I. and C.T. based serial sectioned cadaver specimens has allowed an accurate and highly detailed orientation of the planes of the body and provides the student with a clearer understanding of anatomical structure and pathological anomalies as seen with modern imaging techniques.

COMPARATIVE STUDY OF NMR-CAT IMAGES WITHIN SERIES PLASTINATED SECTIONS

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The complexity of the results from CAT and NMR examinations very often require the aid of normal morphology support to get to certain and undoubted diagnosis. For this purpose, the study and preservation of "slices" from the head of a corpse, after a precise radiography diagram was made.

MATERIAL AND METHODS

In this study, fixed corpses were used. The corpse was frozen at -25°C in order to allow head resection along a

transverse-cervic-thoracic plane. After carefully studying the radiography the following points were found: repere, glabella and opisthocranium, necessary to carry out the first transverse section. Other sections, parallel to the first one were then cut. Each "slice", to either cranial or caudal direction, and was 1 cm thick. After cutting the frozen specimens by means of a circular band saw, the sections were treated to remove any residual organic substance on the cut surface. The sections were then set on blotting-paper to absorb defrosted liquid. A soaked pad with hydrogen peroxide (H₂O₂) was used to liquify the coagulant on the structures. A solution of hydrogen peroxide was also injected into the blood vessels to remove further obstructions. After this treatment, perfectly cleaned sections, from an anatomical (no residual organic substance) and chromatic viewpoint, were obtained, and the surfaces were photographed with a professional camera (LINHOF). At the end of this phase, each section was set on a suitable metallic support to maintain its morphology.

A cylindrical lattice, 50 cm. high was built. Some transverse planes, made up of metal grids, were attached to a steel wire. The planes were 2 cm. apart from each other and were covered by blotting-paper. The sections were at a distance of 4-5 cm. from one another in order to be thoroughly immersed into the substances used in the next phases. Dehydration was by means of acetone at -25°C. The sections were immersed in that solution for 24 hours. The control of the percentage of acetone in the solution follows. If the value of acetone is less than 98%, it will be replaced with new absolute acetone (99%).

The controls were done every 24 hours until acetone remains unchanged (i.e., not absolute acetone) to avoid section shrinking. After this dehydration, forced impregnation with S-10 resin and S-3 hardener start. The lattice was immersed in the resin in the vacuum room and the pump was brought into action. After 5-10 minutes, the resin gradually begins to replace acetone. As this happens, bubbles moving toward the surface of the substance can be observed through the glass sealing the room.

The pump operator should ensure that a constant amount of bubbles be present without pressure changes. In fact, a large quantity of bubbles means that forced impregnation of the sections is too quick. This causes a quantitative change between the amount of acetone going out and the resin replacing it, which causes a shrinking of the organs. Impregnation at room temperature lasts seven days and finishes whenever pressure at 25 mbar doesn't produce any more bubbles. The sections displayed on the lattice grids are taken out of the vacuum to be drained of resin excess present on the surface. Then, the last phase, the cure, begins. For this purpose a slow cure was used. The sections were arranged in two layers in a vessel at a distance of 10 cm. from each other. A small vessel, filled with KOH (Potassium hydroxide), was set on each layer. The potassium hydroxide absorbs humidity therefore catalyzing the hardening reaction of resin. Each day the sections dried because of continuous secretion of small amounts of resin. After one week, when the sections were dry, this phase was complete.